

# Mechanisms Involved in the Respiratory Depressant Actions of Nicotine in Anesthetized Rats

JEWELL W. SLOAN,<sup>1</sup> WILLIAM R. MARTIN AND M. BOSTWICK

*Department of Pharmacology, University of Kentucky, Lexington, KY 40536*

Received 14 November 1988

SLOAN, J. W., W. R. MARTIN AND M. BOSTWICK. *Mechanisms involved in the respiratory depressant actions of nicotine in anesthetized rats.* PHARMACOL BIOCHEM BEHAV 34(3) 559-564, 1989.—In the urethane-pentobarbital anesthetized rat, the respiratory depressant and lethal effects of intravenously infused (–)-nicotine (120 µg/kg/min) or (+)-nicotine (600 µg/kg/min) were effectively prevented by pretreatment with the opioid antagonist, naltrexone, whereas the lethal effect of (–)-nicotine (120 µg/kg/min) was not altered by bilateral adrenalectomy. Further, pretreatment with either the nicotinic ganglion-blocker, mecamylamine, a secondary amine, or the quaternary nicotinic ganglion-blocker, hexamethonium, completely prevented the lethal effects of (–)-nicotine (120 µg/kg/min). These data suggest that central opioidergic and nicotinic processes are involved in nicotine's respiratory depressant and lethal effects.

Nicotine	Opioidergic and nicotinic processes: nicotine lethality	Nicotine's respiratory depressant actions in the rat
Naltrexone prevention of nicotine lethality	Hexamethonium prevention of nicotine lethality	
Mecamylamine prevention of nicotine lethality	Adrenalectomy and nicotine lethality	

BOTH (–)- and (+)-nicotine are known to produce death through respiratory depression (26, 30, 31, 44, 58). Other nicotinic ligands such as cytisine and lobeline also cause death (6,36) due to respiratory depression (13,18). Studies have shown that the respiratory responses to the application of nicotine to the ventrolateral surface of the medulla, its injection intravenously or into the lateral ventricles of the cat, can be prevented by the application of hexamethonium to the ventrolateral medullary surface (16,28). Numerous studies have also demonstrated that nicotinic antagonists such as mecamylamine, which act both centrally and peripherally, alter patterns of nicotine intake in both humans and animals (33).

Since the isolation of endogenous opioid peptides in the brain (32), high concentrations of opiate receptors as well as enkephalins have been found in the solitary nuclei and other medullary areas that affect respiration (4, 5, 8). One of the consequences of acute intoxication by exogenous opioids in man is respiratory depression, a condition which is now routinely treated with narcotic antagonists (43, 46, 47). Endogenous opioids also produce naloxone antagonizable respiratory depression when applied to the ventral surface of the brainstem; when injected into the lateral ventricles and into the pontomedullary subarachnoid space; depression of spontaneous discharge; and depression of L-glutamate induced firing of respiration-related units in the pontine and bulbar respiratory centers (14, 20, 22, 49, 51, 74). Further, it has

recently been shown that nicotine can increase endogenous opioids centrally. After the repeated short-term administration of nicotine (0.1 mg/kg/IP six times at 30-minute intervals) increased levels of methionine enkephalin are found in the rat striatum (52). A number of studies have suggested that opioid mechanisms are involved in the pharmacologic effects of nicotine (12, 34, 53, 54). The purpose of this paper is to explore nicotinic and opioidergic mechanisms in nicotine's respiratory depressant actions in the urethane-pentobarbital-anesthetized rat.

## METHOD

### *Experimental Techniques*

These studies were conducted in the urethane (1 g/kg, IP)-pentobarbital (20 mg/kg/IP) anesthetized female Sprague-Dawley rat (200–300 g). Respiration (inspiratory flow rate) was measured via a tracheal cannula using a Rudolph valve and Gould pneumograph flow transducer. During the course of the experiment, body temperature was maintained at  $37 \pm 1^\circ\text{C}$  by a heat lamp (actuated by a YSI Model 63RC temperature controller connected to the rat by a rectal thermister probe). Minute volume was obtained using a Grass 7P10E polygraph integrator. Tidal volume was calculated from the minute volume and respiratory rate. Respiratory rate and minute volume were recorded continuously on a polygraph (Grass,

<sup>1</sup>Requests for reprints should be addressed to Dr. Jewell W. Sloan, Department of Pharmacology, Room 244, Research Facility #3, University of Kentucky, Lexington, KY 40536.

Model 7D). These measures were also recorded on tape and subsequently digitized (Apple IIe Micro Computer equipped with an ISAAC Data Acquisition and Lab Control Module) for computer analysis.

Drugs were dissolved in normal saline and administered as the free base. (–)- and (+)-nicotine were administered via a cannula inserted into the right external jugular vein at a rate of 0.025 ml/minute for 16 minutes following a 10-minute predrug and a 10-minute saline or drug pretreatment period. The doses of (–)-nicotine (120 µg/kg/min) and (+)-nicotine (600 µg/kg/min) were selected on the basis of dose ranging. These doses were chosen because they reproducibly depressed respiration and caused death in the urethane-pentobarbital anesthetized rat. Doses of hexamethonium, mecamlamine and naltrexone were also chosen by dose ranging. Doses were selected which reproducibly prevented respiratory depression and death in the (–)-nicotine- (120 µg/kg/min) and (+)-nicotine- (600 µg/kg/min) infused rat. Saline, naltrexone, hexamethonium and mecamlamine were administered as an IV bolus over a 1-min period in a volume of approximately 0.16 ml. Residual drug or saline was flushed into the vein with approximately 0.2 ml of saline and observations were made for 10 minutes prior to the administration of nicotine.

#### Data Analysis

The data are presented as the change in response from the appropriate control. The mean change in response for each drug treatment for the different parameters ( $\pm$  S.E.M.) was estimated for each minute and time-response curves were constructed. The data were further analyzed by appropriate unpaired comparisons and by Chi-Square analysis.

#### Drugs and Chemicals

(+)-Nicotine was resolved by Dr. Amy Howell and Dr. W. T. Smith of the University of Kentucky, Department of Chemistry (60). (–)-Nicotine was obtained from Research Plus (Bayone, NJ); hexamethonium bromide, Sigma Chemical Co. (St. Louis, MO); mecamlamine hydrochloride, Dr. Clement Stone, Merck Sharp and Dohme (West Point, PA). Sources of other drugs and chemicals have been identified previously (59).

#### RESULTS

Figure 1 shows that in the urethane-pentobarbital anesthetized rat an IV injection of normal saline had no effect on respiratory measures, whereas naltrexone enhanced tidal volume, but did not alter respiratory rate or minute volume relative to the pretreatment control. Hexamethonium tended to increase all respiratory measures, but these changes were not statistically significant. In contrast, mecamlamine produced a significant enhancement of respiratory rate and minute volume.

The continuous IV infusion of either (–)-nicotine (120 µg/kg/minute) or (+)-nicotine (600 µg/kg/minute) depressed respiratory rate and produced respiratory arrest in 4 of 5 rats. (–)-Nicotine differed from (+)-nicotine in that severe respiratory depression occurred faster and 4 of 5 rats were dead within 4 minutes. Although (+)-nicotine produced some depression of respiratory rate within the first 10 minutes, both minute volume and tidal volume were enhanced during this period. After 10 minutes of infusion, however, the respiratory depressant actions of (+)-nicotine predominated and 4 of 5 rats were dead at 16 minutes (Fig. 2). When rats were pretreated with naltrexone the survival rate was significantly increased after both (–)-nicotine (5 of 6 rats survived,  $\chi^2=4.41$ ,  $p>0.05$ ) and after (+)-nicotine (5 of 5 rats survived,  $\chi^2=6.67$ ,  $p<0.05$ ). In this regard, it can be seen that

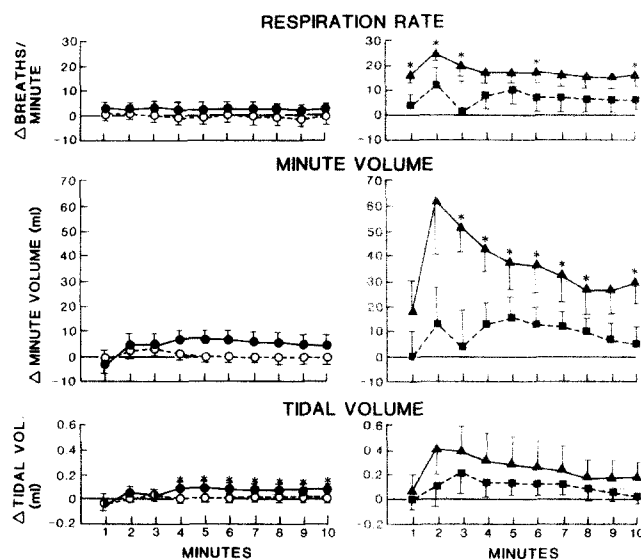


FIG. 1. The effects of naltrexone, saline, mecamlamine and hexamethonium on respiratory rate, tidal volume and minute volume in the urethane-pentobarbital anesthetized rat. Drugs (or saline) were given as an IV bolus in a volume of 1.6 ml of normal saline and flushed into the vein with 0.2 ml of normal saline. Values represent the mean change ( $\pm$  S.E.) from the pretreatment control for the number of animals (shown below) for each minute during the 10-minute observation period. ○: Saline (bolus IV) (n=4); ●: naltrexone (5 mg/kg, bolus IV) (n=6); ■: hexamethonium (1 mg/kg, bolus IV) (n=5); ▲: mecamlamine (1 mg/kg, bolus IV) (n=5). \*Significant change from pretreatment control,  $p<0.05$ .

after naltrexone pretreatment a greater enhancement of minute volume and tidal volume was produced in the presence of (+)- and (–)-nicotine than in their absence (Figs. 1 and 2). Further, this stimulation of respiration in the presence of naltrexone was greater after (–)-nicotine infusion than after (+)-nicotine infusion. Bilateral adrenalectomy did not alter the lethality of (–)-nicotine infusion (120 µg/kg/min). Three of 6 rats survived compared to sham-operated rats where 2 of 4 rats survived (–)-nicotine infusion ( $\chi^2=0$ ).

Respiratory rate and minute volume were significantly enhanced following the infusion of (–)-nicotine (120 µg/kg/min) in both the mecamlamine- and hexamethonium-pretreated rats compared to saline pretreatment and the lethal effects of (–)-nicotine were completely abolished (Fig. 3). It should be pointed out, however, that the changes induced by mecamlamine and hexamethonium after (–)-nicotine were different. Whereas the marked stimulation of respiration induced by mecamlamine was blunted by (–)-nicotine infusion, tidal volume and minute volume were enhanced by (–)-nicotine in the hexamethonium-pretreated rats. This was similar to the effects produced by (–)-nicotine infusion after naltrexone pretreatment (Figs. 1 and 3).

#### DISCUSSION

Although nicotine can stimulate respiration in man (67) and laboratory animals (1, 15, 34, 57), it can also depress respiration (17, 23, 24, 57, 65, 68). These studies present evidence which suggest that nicotine depresses respiration through both nicotinic and opioidergic mechanisms which have a major central component. As previously reported, the urethane-pentobarbital anesthetized rat shows little evidence of the ganglion-stimulant actions of nicotine, while the depressant effects predominate (59,61).

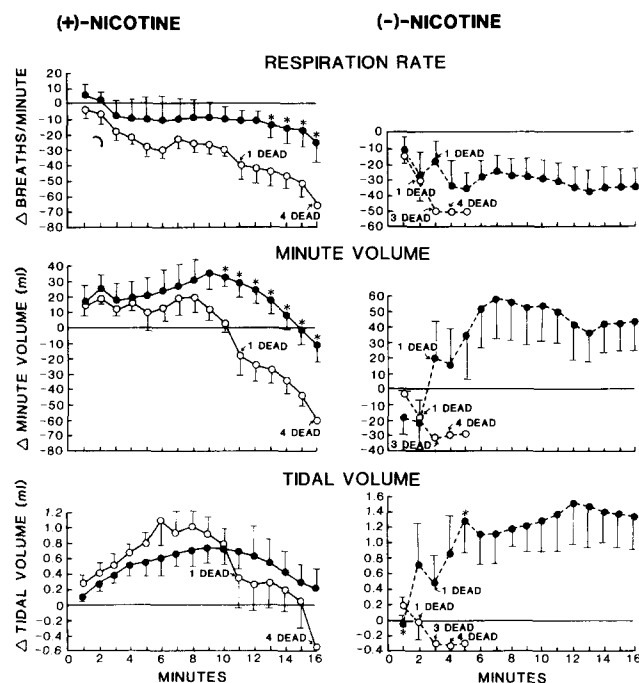


FIG. 2. Comparison of the effects of the continuous IV infusion of (+)-nicotine (600 µg/kg/min, solid lines) and (-)-nicotine (120 µg/kg/min, dashed lines) on respiratory measures and lethality in saline- and naltrexone-pretreated rats. Each point represents the mean change ( $\pm$  S.E.) from the appropriate pretreatment condition for each minute for the number of rats indicated below in parentheses. ○-○: Saline pretreatment (n=5); ●-●: naltrexone pretreatment (5 mg/kg IV) (n=5); ○-○: saline pretreatment (n=5); ●-●: naltrexone pretreatment (5 mg/kg IV) (n=6). \*Different from saline pretreatment,  $p < 0.05$ .

The effects of naltrexone and other opioid antagonists on respiration are complex and may be the consequence of several modes of action. It has generally been reported that both naltrexone and naloxone are devoid of significant effects on respiration in healthy men and animals, but have been shown to play a role in altering respiration brought about by several pathologic conditions, presumably by antagonizing endogenous opioids (27, 43, 47). In this regard, naltrexone has been shown to partially reverse the depressed level of consciousness and the depressed flexor reflex response, but not respiration in the pentobarbital anesthetized chronic spinal dog. These data were interpreted as suggesting that some of naltrexone's actions are mediated through  $\kappa$  receptors where the effects of pentobarbital are antagonized directly, or, alternatively, indirectly by reversing the effects of pentobarbital-released endogenous substances (25). In the present experiments, naltrexone alone produced a slight enhancement of respiratory measures in the urethane-pentobarbital anesthetized rat. The ability of naltrexone to stimulate respiration may be related to the level of endogenous opioid tone. In the present acute studies endogenous opiates may have been released in response to the surgical procedures, whereas in the chronic spinal dog studies (25) the animals had recovered from surgery.

Naltrexone pretreatment prevented the lethal effects of both (-)- and (+)-nicotine suggesting the involvement of opioidergic processes in nicotine's respiratory depressant effects. These data are consistent with evidence obtained in man which have demonstrated that naloxone pretreatment reversed postsmoking respiratory depression (67). It has been shown that nicotine can release opioid peptides from the perfused dog and bovine adrenal gland and from adrenal medulla chromaffin cells in primary culture (19,

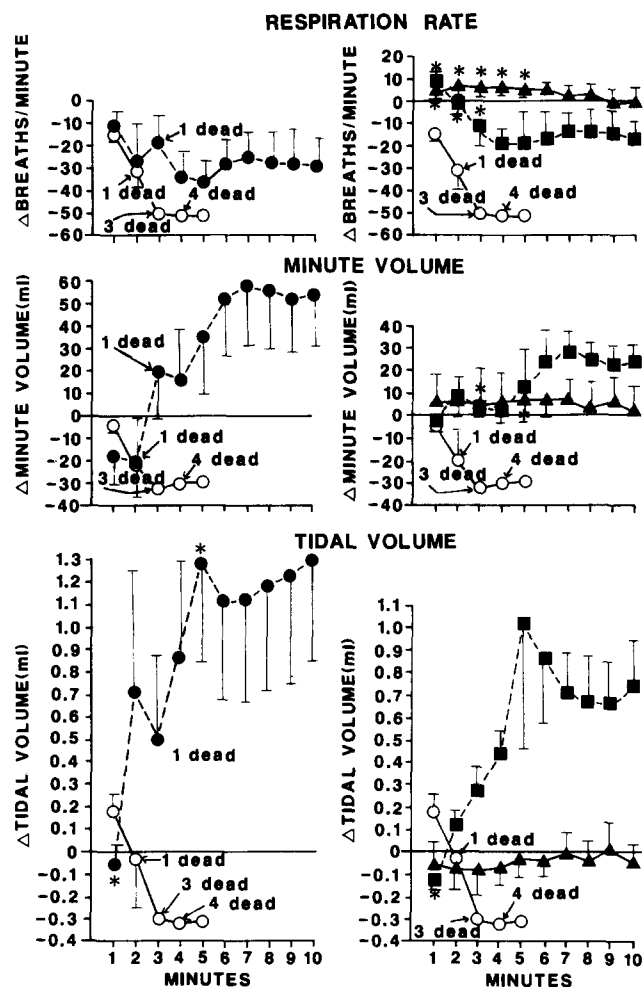


FIG. 3. Comparison of the effects of the continuous IV infusion of (-)-nicotine (120 µg/kg/min) on lethality and respiratory measures in the saline-, naltrexone-, hexamethonium- and mecamylamine-pretreated anesthetized rat. Each point represents the mean change ( $\pm$  S.E.) from the appropriate pretreatment condition for each minute for the number of experiments indicated. ○: Saline pretreatment (n=5); ●: naltrexone pretreatment (5 mg/kg) (n=6); ■: hexamethonium pretreatment (1 mg/kg) (n=5); ▲: mecamylamine pretreatment (1 mg/kg) (n=5). \*Different from saline pretreatment,  $p < 0.05$ .

35, 70, 72). If the release of opioid peptides from the adrenal gland by nicotine is responsible for its respiratory depressant actions, then adrenalectomy should abolish this effect. The adrenal gland does not appear to be the essential component involved in nicotine's respiratory depressant effects since bilateral adrenalectomy did not alter the lethality of nicotine in the anesthetized rat. This observation argues against a peripheral opioidergic site for nicotine's respiratory depressant effects. Endogenous opioids and opioid receptors are found in the medulla of the rat (4) as are nicotine binding sites (38,45) in areas closely associated with central respiratory control in the rat (29). Moreover, it has been demonstrated that after intracisternal administration in dogs, beta-endorphin induced a naloxone-reversible respiratory depressant action (49). Met-enkephalin applied to the ventral surface of the cat brainstem depressed both tidal volume and respiratory rate (21). Thus, there is substantive data indicating the existence of a medullary opioid respiratory depressant mechanism.

Both mecamlamine and hexamethonium tended to enhance respiration, although this was a statistically significant effect for mecamlamine only. Hexamethonium, however, has been reported previously to produce an initial increase in tidal volume when applied to the chemoreceptive zones of the medulla oblongata of the chloralose-urethane anesthetized cat (16). The mechanism by which mecamlamine and hexamethonium enhance respiration is unknown. One possibility with regard to mecamlamine is that this effect may be related to mecamlamine's ability to stimulate presynaptic nicotine receptors to release endogenous substances that in turn stimulate respiration (40). Another hypothesis is that both mecamlamine and hexamethonium are antagonizing endogenous nicotinic tone. Mecamlamine is a secondary amine that penetrates the blood-brain barrier (7, 9, 48) and acts both pre- and postsynaptically (56). It has been shown to have selectivity in antagonizing both the peripheral and central effects of nicotine (10).

Hexamethonium is a quaternary ammonium ganglion-blocking agent that acts only at postsynaptic nicotinic receptors (10) and is generally thought to penetrate the blood-brain barrier poorly and, thus, its blocking actions have been used to distinguish the peripheral from the central actions of nicotine (11, 50, 62, 63). Other investigators, however, have found that hexamethonium does enter the brain in pharmacologically significant amounts (3,41). Further, many of the central effects of systemically administered nicotine such as antinociception in the rat (69); convulsions in the rat and mouse (39, 64, 73); emesis in the cat (37); the prostration immobilization syndrome induced by centrally administered nicotine in the rat (56); the ventilatory response to nicotine microinjected into the third ventricle of the cat (42); the response to its intravenous administration in the dog, cat and rabbit (17); and the discrimination of intracerebroventricularly adminis-

tered nicotine in the rat (55) can be prevented by either systemically or centrally administered hexamethonium.

Although hexamethonium and mecamlamine differed in their effects on nicotine-induced respiratory depression in that hexamethonium-treated rats showed more nicotine-induced respiratory stimulation, both drugs completely prevented the lethal effects of (-)-nicotine in the urethane-pentobarbital anesthetized rat. The respiratory stimulant actions of mecamlamine, hexamethonium and naltrexone may be due to their respective abilities to antagonize endogenous nicotinic cholinergic and opioidergic tone in the respiratory centers. In this regard, mecamlamine was more effective than hexamethonium and naltrexone. In the presence of either hexamethonium, mecamlamine or naltrexone, (-)-nicotine produced an increase in tidal volume and minute volume. This stimulatory effect of (-)-nicotine was greater in hexamethonium- and naltrexone-pretreated rats than in mecamlamine-pretreated rats. These observations can be reconciled with the hypothesis that these drugs are blocking endogenous and exogenous nicotinic- and opioidergic-mediated respiratory depression. Further, the data are entirely consistent with the idea that this blockade is mediated by competitive antagonism of endogenous and exogenous agonists. The studies, however, were not designed to address this issue.

These data taken together suggest that central sites are involved in nicotine's lethal effects and support the concept that central opioidergic and nicotinic processes probably act in concert to modulate nicotine's effects on respiration. Further studies are necessary, however, to locate the possible sites where this modulation occurs.

#### ACKNOWLEDGEMENT

This research was supported by a grant from the Tobacco and Health Research Institute of the University of Kentucky.

#### REFERENCES

1. Abood, L. G.; Lowy, K.; Tometsko, A.; MacNeil, M. Evidence for a noncholinergic site for nicotine's action in brain: Psychopharmacological, electrophysiological and receptor binding studies. *Arch. Int. Pharmacodyn. Ther.* 237(2):213-229; 1979.
2. Abood, L. G.; Reynolds, D. T.; Booth, H.; Bidlack, J. M. Sites and mechanisms for nicotine's action in the brain. *Neurosci. Biobehav. Rev.* 5:479-486; 1981.
3. Asghar, K.; Roth, L. J. Entry and distribution of hexamethonium in the central nervous system. *Biochem. Pharmacol.* 20:2787-2795; 1971.
4. Atweh, S. F.; Kuhar, M. J. Autoradiographic localization of opiate receptors in rat brain. I. Spinal cord and lower medulla. *Brain Res.* 124:53-67; 1977.
5. Atweh, S. F.; Kuhar, M. J. Autoradiographic localization of opiate receptors in rat brain. II. The brain stem. *Brain Res.* 129:1-12; 1977.
6. Barlow, R. B.; McLeod, L. J. Some studies on cytosine and its methylated derivatives. *Br. J. Pharmacol.* 35:161-174; 1969.
7. Bennett, G.; Tyler, C. Mecamlamine and its mode of action. *Lancet* II:218-222; 1957.
8. Berger, A. J.; Mitchell, R. A.; Severinghaus, J. W. Regulation of respiration (Parts I-III) *N. Engl. J. Med.* 297:92-97, 138-143, 194-201; 1977.
9. Brodie, B. B.; Kurz, H.; Schanker, L. S. The importance of dissociation constant and lipid-solubility in influencing the passage of drugs into the cerebrospinal fluid. *J. Pharmacol. Exp. Ther.* 130: 20-35; 1960.
10. Brown, D. A. Locus and mechanism of action of ganglion-blocking agents. In: Born, G. V. R.; Farah, A.; Herken, H.; Welch, A. D., eds. *Handbook of experimental pharmacology*, vol. 53. Berlin: Springer-Verlag; 1980.
11. Clark, P. B. S.; Kumar, R. The effects of nicotine on locomotor activity in non-tolerant and tolerant rats. *Br. J. Pharmacol.* 78: 329-337; 1983.
12. Corrigan, W. A.; Herling, S.; Coen, K. M. Evidence for opioid mechanisms in the behavioral effects of nicotine. *Psychopharmacology* (Berlin) 96:29-35; 1988.
13. Dale, H. H.; Laidlaw, P. P. The physiological action of cytosine, the active alkaloid of laburnum (*Cytisus laburnum*). *J. Pharmacol. Exp. Ther.* 3:205-221; 1912.
14. Denavit-Saubie, M.; Champagnat, J.; Zieglans-Berger, W. Effects of opiates and methionine-enkephalin on pontine and bulbar respiratory neurones in the cat. *Brain Res.* 138:585-590; 1978.
15. Dev, N. B.; Loeschcke, H. H. Topography of the respiratory and circulatory responses to acetylcholine and nicotine on the ventral surface of the medulla oblongata. *Pflugers Arch.* 379:19-27; 1979.
16. Dev, N. B.; Loeschcke, H. H. A cholinergic mechanism involved in the respiratory chemosensitivity of the medulla oblongata in the cat. *Pflugers Arch.* 379:29-36; 1979.
17. Domaye, A. On the action of nicotine upon respiration with particular reference to its blockade by hexamethonium. *Jpn. J. Pharmacol.* 5:1-10; 1955.
18. Edmunds, C. W. On the action of lobeline. *Am. J. Physiol.* 11:79-102; 1904.
19. Eiden, L. E.; Giraud, P.; Dave, J. R.; Hotchkiss, A. J.; Affolter, H. U. Nicotinic receptor stimulation activates enkephalin release and biosynthesis in adrenal chromaffin cells. *Nature* 312:662-663; 1984.
20. Florez, J.; Hurler, M. A.; Mediavilla, A. Respiratory responses to opiates applied to the medullary ventral surface. *Life Sci.* 31: 2189-2192; 1982.
21. Florez, J.; Mediavilla, A. Respiratory and cardiovascular effects of met-enkephalin applied to the ventral surface of the brain stem. *Brain Res.* 138:585-590; 1977.

22. Florez, J.; Mediavilla, A.; Pazos, A. Respiratory effects of  $\beta$ -endorphine, D-Ala<sup>2</sup>-met-enkephalinamide and met-enkephalin injected into the lateral ventricle and the pontomedullary subarachnoid space. *Brain Res.* 199:197-206; 1980.
23. Franke, F. E. Respiratory failure in acute nicotine poisoning. *J. Physiol.* 123:69-70; 1938.
24. Franke, F. E.; Thomas, J. E. A study of the cause of death in experimental nicotine poisoning in dogs. *J. Pharmacol.* 48:199-208; 1933.
25. Gilbert, P. E.; Martin, W. R. Antagonism of the effects of pentobarbital in the chronic spinal dog by naltrexone. *Life Sci.* 20:1401-1406; 1977.
26. Gold, H.; Brown, F. A contribution to the pharmacology of nicotine. *J. Pharmacol.* 54:463-476; 1935.
27. Gonzalez, J. P.; Brogden, R. N. Naltrexone: A review of its pharmacodynamic and pharmacokinetic properties and therapeutic efficacy in the management of opioid dependence. *Drugs* 35:192-213; 1988.
28. Haxhius, M. A.; Van Lunteren, E.; Van DeGraff, W. B.; Strohl, K. P.; Bruce, E. N.; Mitra, J.; Cherniack, N. S. Actions of nicotine on the respiratory activity of the diaphragm and geniglossus muscles and the nerves that innervate them. *Respir. Physiol.* 57:153-169; 1984.
29. Hedner, J. Neuropharmacological aspects of central respiratory regulation. *Acta Physiol. Scand.* [Suppl.] 524; 1983.
30. Hicks, C. S.; Mackay, M. E.; Sinclair, D. A. The comparative pharmacology of the nor-nicotines. *Aust. J. Exp. Biol. Med. Sci.* 25:363-372; 1947.
31. Hicks, C. S.; Sinclair, D. A. Toxicities of the optical isomers of nicotine and normicotine. *Aust. J. Exp. Biol. Med. Sci.* 25:83-86; 1947.
32. Hughes, J.; Smith, T. W.; Kosterlitz, H. W.; Fothergill, L. A.; Morgan, B. A.; Morris, H. R. Identification of two related pentapeptides from the brain with potent opiate agonist activity. *Nature* 258:577-579; 1975.
33. Jarvick, M. W.; Henningfield, J. E. Pharmacological treatment of tobacco dependence. *Pharmacol. Biochem. Behav.* 30:279-294; 1988.
34. Kamerling, S. G.; Wettstein, J. G.; Sloan, J. W.; Su, T. P.; Martin, W. R. Interaction between nicotine and endogenous opioid mechanisms in the unanesthetized dog. *Pharmacol. Biochem. Behav.* 17:733-740; 1982.
35. Kilpatrick, D. L.; Lewis, R. V.; Stein, S.; Udenfriend, S. Release of enkephalins and enkephalin-containing polypeptides from perfused beef adrenal glands. *Proc. Natl. Acad. Sci. USA* 77:7473-7475; 1980.
36. Klocking, H. P.; Damm, G.; Richter, M. The influence of drugs on the acute toxicity of cytosine. *Arch. Toxicol. Suppl.* 4:402-404; 1980.
37. Laffan, R. J.; Borison, H. L. Emetic action of nicotine and lobeline. *J. Pharmacol. Exp. Ther.* 121:468-476; 1957.
38. Larsson, C. Nicotinic receptors in the central nervous system: Methodological and functional aspects. *Acta University Upsaliensis*, 1985.
39. Laurence, D. R.; Stacey, R. S. The effect of methonium compounds on nicotine convulsions. *Br. J. Pharmacol.* 7:80-84; 1952.
40. Lees, G. M.; Nishi, S. Analysis of the mechanism of action of some ganglion-blocking drugs in the rabbit superior cervical ganglion. *Br. J. Pharmacol.* 46:78-88; 1972.
41. Levine, R. R. Presence of certain onium compounds in brain tissue following intravenous administration to rats. *Nature* 184:1412-1414; 1959.
42. McCarthy, L. E.; Borison, H. L. Separation of central effects of CO<sub>2</sub> and nicotine on ventilation and blood pressure. *Respir. Physiol.* 15:321-330; 1972.
43. McNicholas, L. F.; Martin, W. R. New and therapeutic roles for naloxone and related opioid antagonists. *Drugs* 27:81-93; 1984.
44. Macht, D. I.; Davis, M. E. Toxicity of alpha- and beta-nicotines and normicotines: An inquiry into chemopharmacodynamic relationships. *J. Pharmacol. Exp. Ther.* 1(1):93-99; 1934.
45. Martin, B. R.; Aceto, M. D. Nicotine binding sites and their localization in the central nervous system. *Neurosci. Biobehav. Rev.* 6:473-478; 1981.
46. Martin, W. R. Opioid antagonists. *Pharmacol. Rev.* 19:463-521; 1967.
47. Martin, W. R. Pharmacology of opioids. *Pharmacol. Rev.* 35:283-323; 1984.
48. Milne, M. D.; Rowe, G. G.; Somers, K.; Muehrcke, R. C.; Crawford, M. A. Observations on the pharmacology of mecamlamine. *Clin. Sci.* 16:599-614; 1957.
49. Moss, I. R.; Friedman, E. Beta-endorphin: Effects on respiratory regulation. *Life Sci.* 23:1271-1276; 1978.
50. Paton, W. D. M.; Zaimis, E. J. The methonium compounds. *Pharmacol. Rev.* 4:219-253; 1952.
51. Paxos, A.; Florez, J. Interaction of naloxone with  $\mu$ - and  $\delta$ -opioid agonists on the respiration of rats. *Eur. J. Pharmacol.* 87:309-314; 1983.
52. Pierzchala, K.; Houdi, A. A.; Van Loon, G. R. Nicotine-induced alterations in brain regional concentrations of native and cryptic Met- and Leu-enkephalin. *Peptides* 8:1035-1043; 1987.
53. Pomerleau, O. F.; Fertig, J. B.; Seyler, L. E.; Jaffe, J. Neuroendocrine reactivity to nicotine in smokers. *Psychopharmacology (Berlin)* 81:61-67; 1983.
54. Pomerleau, O. F.; Pomerleau, C. S. Neuroregulators and the reinforcement of smoking: Towards a biobehavioral explanation. *Neurosci. Biobehav. Rev.* 8:503-513; 1984.
55. Romano, C.; Goldstein, A.; Jewell, N. P. Characterization of the receptor mediating the nicotine discriminative stimulus. *Psychopharmacology (Berlin)* 74:310-315; 1981.
56. Schwab, L. S.; Kritzer, M. F. The effect of cholinergic antagonists on a central response to nicotine. *Experientia* 38:119-120; 1982.
57. Silvette, H.; Hoff, E. C.; Larson, P. S.; Haag, H. B. The actions of nicotine on central nervous system functions. *Pharmacol. Rev.* 14:137-173; 1962.
58. Sloan, J. W.; Martin, W. R.; Bostwick, M. Cardiovascular and respiratory effects of (+) and (-)-nicotine in the anesthetized rat. *Pharmacology* 28(3):156; 1986.
59. Sloan, J. W.; Martin, W. R.; Bostwick, M.; Hook, R.; Wala, E. The comparative binding characteristics of nicotinic ligands and their pharmacology. *Pharmacol. Biochem. Behav.* 30:255-267; 1988.
60. Sloan, J. W.; Martin, W. R.; Hernandez, J.; Hook, R. Binding characteristics of (-)- and (+)-nicotine to the rat brain P<sub>2</sub> fraction. *Pharmacol. Biochem. Behav.* 23:987-993; 1985.
61. Sloan, J. W.; Martin, W. R.; Smith, W. T. Multiple nicotinic receptors: nicotinic ligands with different specificities. In: Martin, W. R.; Van Loon, G. R.; Iwamoto, E. T.; Davis, L., eds. *Tobacco smoking and nicotine: A neurobiological approach*. New York: Plenum Press; 1987:451-465.
62. Spealman, R. D.; Goldberg, S. R.; Gardner, M. L. Behavioral effects of nicotine: Schedule-controlled responding by squirrel monkeys. *J. Pharmacol. Exp. Ther.* 216(3):484-491; 1981.
63. Stoleran, I. P.; Kuman, R.; Pratt, J. A.; Reavill, C. Discriminative stimulus effects of nicotine: correlation with binding studies. In: Martin, W. R.; Van Loon, G. R.; Iwamoto, E. T.; Davis, L., eds. *Advances in behavioral biology 31: Tobacco smoking and nicotine: A neurobiological approach*. New York: Plenum Publishing Co.; 1987:113-124.
64. Stone, C. A.; Mecklenburg, K. L.; Torchiana, M. L. Antagonism of nicotine-induced convulsions by ganglionic blocking agents. *Arch. Int. Pharmacodyn.* 127:419-434; 1958.
65. Thomas, J. E.; Franke, F. E. Peripheral paralysis of a cause of respiratory failure in acute nicotine poisoning. *J. Pharmacol.* 23:150-151; 1924.
66. Thomas, J. E.; Franke, F. E. The site of the toxic action of nicotine on the respiratory mechanism. *J. Pharmacol.* 34:111-135; 1928.
67. Tobin, M. J.; Jenouri, G.; Sackner, M. A. Effect of naloxone on change in breathing pattern with smoking. *Chest* 82:530-537; 1982.
68. Tobin, M. J.; Schneider, A. W.; Sackner, M. A. Breathing pattern during and after smoking cigarettes. *Clin. Sci.* 63:473-483; 1982.
69. Tripathi, H. L.; Martin, B. R.; Mario, D. A. Nicotine-induced antinociception in rats and mice: correlation with nicotine brain levels. *J. Pharmacol. Exp. Ther.* 221(1):91-96; 1982.
70. Viveros, O. H.; Diliberto, E. J., Jr.; Hazum, E.; Chang, K. J. Enkephalins as possible adrenomedullary hormones: Storage, secretion and regulation of synthesis. In: Costa, E.; Trabucchi, M., eds. *Neural peptides and neuronal communication*. New York: Raven Press; 1980.
71. Viveros, O. H.; Diliberto, E. J., Jr.; Hazum, E.; Chang, K. J. Opiate-like materials in the adrenal medulla: Evidence for storage and secretion with catecholamines. *Mol. Pharmacol.* 16:1101-1108; 1979.

72. Wilson, S. P.; Chang, K. J.; Viveros, O. H. Proportional secretion of opioid peptides and catecholamines from adrenal chromaffin cells in culture. *J. Neurosci.* 2:1150–1156; 1982.
73. Yamamoto, I.; Otori, K.; Inoki, R. Pharmacological studies on antagonists against nicotine-induced convulsions and death. *Jpn. J. Pharmacol.* 16:402–415; 1966.
74. Zobrist, R. H.; Allerton, H. W.; Isom, G. E. Characterization of the respiratory activity of (D-ala<sup>2</sup>) methionine-enkephalinamide. *Eur. J. Pharmacol.* 770:121–128; 1981.